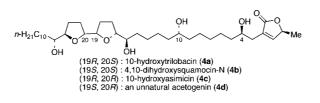
Highly Stereoselective and Modular Syntheses of 10-Hydroxytrilobacin and Three Diastereomers via Stereodivergent [3 + 2]-Annulation Reactions

Chan Woo Huh and William R. Roush*

Department of Chemistry, Scripps Florida, Jupiter, Florida 33458 roush@scripps.edu

Received June 2, 2008

ABSTRACT



A convergent synthesis of the annonaceous acetogenin, 10-hydroxytrilobacin (4a), was accomplished by using the [3 + 2]-annulation reaction of tetrahydrofuranyl carboxaldehyde 2a and allylsilane 3. The stereodivergency of the [3 + 2]-annulation reaction made it possible to achieve modular, highly stereoselective syntheses of three 10-hydroxytrilobacin diastereomers from the same precursors by using simple modifications of reaction conditions.

The annonaceous acetogenins are fatty acid derived natural products isolated from *Annonaceae* species (custard apple).¹ As of 2005, more than 400 acetogenins had been reported.^{1a} The acetogenins are characterized by their ability to inhibit mitochondria complex I of the mammalian and insect mitochondrial electron transport system.^{1a}

Among the numerous natural acetogenins, the stereochemically diverse group of adjacent bis-THF acetogenins (especially those with hydroxyl groups flanking both ends of the bis-THF unit) have attracted the attention of many synthetic laboratories² because of their significant biological activity against various human cancer cell lines.^{1b}

While it has been stated in a review that the stereochemistry of the bis-THF core is not as important as the number of THF units and the number of hydroxyl groups in determining the biological potency of the various acetogenins,^{1b} it is readily apparent from comparison of the activity of diastereomeric bis-THF acetogenins that the stereochemistry of the bis-THF unit contributes substantially to their biological profiles. For example, the LC₅₀'s for bullatacin and asimicin (which differ at a single stereocenter) against MCF-7 are $<10^{-12} \mu g/mL$ and $8.5 \times 10^{-1} \mu g/mL$, respectively.³ However, the factors that govern the relationship of acetogenin stereochemistry to biological activity are unknown at present. Thus, generation of all possible diastereomers of the bis-THF acetogenins would enable this question to be addressed. Development of a modular and stereochemically general synthesis of adjacent bis-THF acetogenins amenable to SAR studies remains a major

ORGANIC

For recent reviews: (a) Bermejo, A.; Figadère, B.; Zafra-Poloa, M.-C.; Barrachina, I.; Estornell, E.; Cortes, D. *Nat. Prod. Rep.* 2005, *22*, 269.
 (b) Alali, F. Q.; Liu, X.-X.; McLaughlin, J. L. *J. Nat. Prod.* 1999, *62*, 504.
 (c) Zafra-Polo, M. C.; Figadere, B.; Gallardo, T.; Tormo, J. R.; Cortes, D. *Phytochemistry* 1998, *48*, 1087.

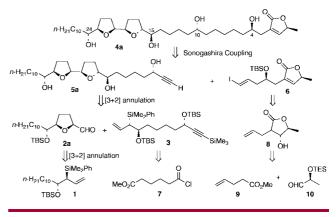
^{(2) (}a) Tinsley, J. M.; Mertz, E.; Chong, P. Y.; Rarig, R. A.; Roush, W. R. Org. Lett. 2005, 7, 4245. (b) Tinsley, J. M.; Roush, W. R. J. Am. Chem. Soc. 2005, 127, 10818. (c) Narayan, R. S.; Borhan, B. J. Org. Chem. 2006, 71, 1416. (d) Keum, G.; Hwang, C. H.; Kang, S. B.; Kim, Y.; Lee, E. J. Am. Chem. Soc. 2005, 127, 10396. (e) Nattrass, G. L.; Di'ez, E.; McLachlan, M. M.; Dixon, D. J.; Ley, S. V. Angew. Chem., Int. Ed. 2005, 44, 580. (f) Head, G. D.; Whittingham, W. G.; Brown, R. C. D. Synlett 2004, 1437. (g) Marshall, J. A.; Piettre, A.; Paige, M. A.; Valeriote, F. J. Org. Chem. 2003, 68, 1780. (h) Emde, U.; Koert, U. Eur. J. Org. Chem. 2000, 2000, 1889.

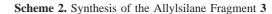
⁽³⁾ He, K.; Shi, G.; Zhao, G.-X.; Zeng, L.; Ye, Q.; Schwedler, J. T.; Wood, K. V.; McLaughlin, J. L. J. Nat. Prod. **1996**, 59, 1029.

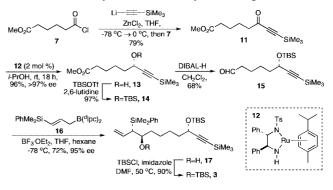
challenge.⁴ Current approaches to the synthesis of libraries of bis-THF acetogenin analogues require use of multiple complex starting materials⁵ or additional steps to install flanking hydroxyl groups after establishment of the bis-THF core.⁶

Preliminary studies in our laboratory demonstrated the potential for use of the [3 + 2]-annulation reaction⁷ for synthesis of adjacent bis-THF acetogenin analogues.⁸ The stereochemistry of the two THF rings is set by using either a nonchelate (BF₃•OEt₂ catalyzed) or a chelate controlled (SnCl₄ catalyzed) [3 + 2]-annulation reaction of a chiral allylsilane with the appropriate aldehyde coupling partner. Although we previously reported syntheses of asimicin and bullatacin using the [3 + 2]-annulation reaction,^{2a,b} these syntheses were too linear and correspondingly lengthy. This prevented us from using these routes to capitalize on the implicit stereodivergency of the [3 + 2]-annulation reaction for generation of stereochemically diverse bis-THF acetogenin analogues. Therefore, we have developed and report herein a convergent, highly stereoselective and stereodivergent synthesis of four representative adjacent bis-THF acetogenins using [3 + 2]-annulation reactions of chiral, nonracemic β -alkoxy allylsilanes 1 and 3.

We targeted the synthesis of 10-hydroxytrilobacin³ (4a), a natural acetogenin with a cis/erythro/trans bis-THF core unit which displays high cytotoxicity toward two human cancer cell lines, A-549 (LC₅₀ = $1.0 \times 10^{-8} \mu \text{g/mL}$) and MCF-7 (LC₅₀ = $1.9 \times 10^{-8} \,\mu \text{g/mL}$).³ We anticipated 10hydroxytrilobacin (4a) could be generated by Sonogashira coupling⁹ of the bis-THF alkyne fragment 5a and the butenolide-containing vinyl iodide 6 (Scheme 1). The bis-THF unit of 5a would be accessed by a nonchelate-controlled [3 + 2]-annulation of *cis*-THF aldehyde **2a** and allylsilane 3. On the basis of earlier studies, it was anticipated that this [3+2]-annulation would be stereochemically mismatched⁸ and therefore synthetically challenging. The cis-THF aldehyde 2a would be derived from allylsilane 1 by a nonchelatecontrolled [3 + 2]-annulation.^{7c} The allylsilane subunit of **3** would be introduced by using an asymmetric γ -silvlallylboration reaction,¹⁰ whereas the chiral propargyl alcohol center of 3 would be generated by transfer hydrogenation of a propargyl ketone precursor.¹¹ We anticipated that the butenoScheme 1. Retrosynthetic Analysis of 10-Hydroxytrilobacin (4a)







lide 6 could be obtained from the β -hydroxy- γ -lactone 8, which in turn would be assembled via the aldol reaction of ester 9 and aldehyde 10.

The synthesis of allylsilane **3** began with nucleophilic addition of trimethylsilylethynyl lithium to methyl adipoyl chloride (**7**),¹² followed by a Noyori asymmetric transfer hydrogenation using chiral catalyst **12**¹³ which provided propargyl alcohol **13** with 97% ee and 76% yield for two steps (Scheme 2). After protection of **13** as the TBS ether (97%), the ester unit of **14** was reduced by treatment with DIBAL-H to yield aldehyde **15** (68%). Asymmetric allylation of **15** with γ -silylallylborane **16**¹⁰ and then protection of the new hydroxyl group as a TBS ether afforded **3** in 65% yield for the two steps, and with 95% ee.

The synthesis of the butenolide fragment **6** was initiated by *B*-iododicyclohexylborane-mediated aldol reaction¹⁴ of ester **9** and chiral aldehyde **10**,¹⁵ followed by lactonization triggered by treatment of the aldol mixture with HF (Scheme 3). Lactone **8** was obtained as the major component of a mixture of diastereomers (4.7:1, ratio of **8** to the sum of three

^{(4) (}a) Kojima, N.; Maezaki, N.; Tominaga, H.; Yanai, M.; Urabe, D.; Tanaka, T. *Chem.-Eur. J.* **2004**, *10*, 672. For library approaches to mono-THF acetogenins: (b) Kojima, N.; Maezaki, N.; Tominaga, H.; Asai, M.; Yanai, M.; Tanaka, T. *Chem.-Eur. J.* **2003**, *9*, 4980. (c) Curran, D. P.; Zhang, Q.; Richard, C.; Lu, H.; Gudipati, V.; Wilcox, C. S. J. Am. Chem. Soc. **2006**, *128*, 9561.

⁽⁵⁾ Das, S.; Li, L. S.; Abraham, S.; Chen, Z.; Sinha, S. C. J. Org. Chem. 2005, 70, 5922.

⁽⁶⁾ Wysocki, L. M.; Dodge, M. W.; Voight, E. A.; Burke, S. D. Org. Lett. 2006, 8, 5637.

^{(7) (}a) Masse, C. E.; Panek, J. S. *Chem. Rev.* **1995**, *95*, 1293. (b) Panek, J. S.; Beresis, R. *J. Org. Chem.* **1993**, *58*, 809. (c) Micalizio, G. C.; Roush, W. R. *Org. Lett.* **2000**, *2*, 461.

⁽⁸⁾ Mertz, E.; Tinsley, J. M.; Roush, W. R. J. Org. Chem. 2005, 70, 8035.

⁽⁹⁾ Sonogashira, K.; Tohda, Y.; Hagihara, N. Tetrahedron Lett. 1975, 16, 4467.

⁽¹⁰⁾ Roush, W. R.; Pinchuk, A. N.; Micalizio, G. C. Tetrahedron Lett. 2000, 41, 9413.

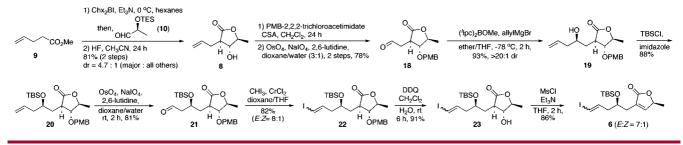
⁽¹¹⁾ Matsumura, K.; Hashiguchi, S.; Ikariya, T.; Noyori, R. J. Am. Chem. Soc. 1997, 119, 8738.

⁽¹²⁾ Carbery, D. R.; Reignier, S.; Myatt, J. W.; Miller, N. D.; Harrity, J. P. A. Angew. Chem., Int. Ed. 2002, 41, 2584.

⁽¹³⁾ Haack, K.-J.; Hashiguchi, S.; Fujii, A.; Ikariya, T.; Noyori, R. Angew. Chem., Int. Ed. 1997, 36, 285.

⁽¹⁴⁾ Ganesan, K.; Brown, H. C. J. Org. Chem. 1994, 59, 2336.
(15) Bongini, A.; Camerini, R.; Hofman, S.; Panunzio, M. Tetrahedron Lett. 1994, 35, 8045.

Scheme 3. Synthesis of Vinyl Iodide Fragment 6



Scheme 4. Stereodivergent Syntheses of Bis-THF Fragments 25a-d 1) TBSOTF nonchelate SiMe₂Ph 2,6-lutidine 2) Pd(OH)₂, H₂ mismatched control OTBS n-H21C10 3) SO3 pyr, DMSO 3, SnCl₄ nonchelate control OTBS i-Pr2NEt, CH2Cl2 твѕо CH2Cl2 4 Å MS, -78 °C твѕо SiMeg 25a 89% (3 steps) 77%, 15:1 dr 22 1) BnOCH₂CHO óн chelate control SiMe₂Ph BF3 OEt2, -78 °C 1) Ac₂O, pyridine 2) Pd(OH)₂, H₂ mismatched OTBS 24a 2) TBAE 3H₀O n-H21C 79% (2 steps) 3) SO3 pyr, DMSO 3. SnCl CH₂Cl₂, 4 Å MS, -78 °C AcÓ **Ö**TBS Ĥ AcÒ SiMe₂Ph i-Pr2NEt, CH2Cl2 SiMe 52%, 20:1 dr 25t 66% (3 steps) 2b n-H21C10 о́твs chelate SiMe₂Ph matcheo control OTBS 1) BnOCH₂CHO n-H21C10 3, SnCl₄ 1) TBSOTE SnCl₄, -78 °C 2.6-lutidine 2) TBAF 3H₂O CH2Cl2.4 Å MS, 0 °C TBSO **OTBS** SiMe: 2) Pd(OH)₂, H₂ 25c 67%. >20:1 dr 62% (2 steps n-H₂₁C 3) SO3 pyr, DMSO chelate control i-ProNEt, CHoClo óн твѕо nonchelate SiMe₂Ph matched 73% (3 steps) OTBS control 24t 2c n-H₂ 3, BF3 OEt2 CH2Cl2, 4 Å MS, -78 °C **OTBS** твѕо `SiMe₃ 67%, >20:1 dr 25d

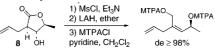
minor isomers, 81% combined yield). The β -hydroxyl group of **8** was protected as a *p*-methoxybenzyl ether (PMB) by using *p*-methoxylbenzyl 2,2,2-trichloroacetimidate and catalytic CSA.¹⁶ One-pot oxidative cleavage¹⁷ of the terminal vinyl group then afforded aldehyde **18** in 78% yield for the two steps. Asymmetric allylation¹⁸ of **18** provided allylation product **19** with >20:1 ds (93% yield). Homoallylic alcohol **19** was then protected as a TBS ether, and the terminal vinyl group was subsequently oxidatively cleaved¹⁷ to furnish aldehyde **21** (81% yield). The vinyl iodide was then installed under modified Takai alkenylation conditions,¹⁹ and the PMB ether was removed using DDQ to give alcohol **23** (74% for two steps). Finally, β -elimination of the hydroxyl group of **23** by treatment with MsCl and Et₃N afforded butenolide fragment **6** (86%).²⁰

The bis-THF fragment **25a**, needed for synthesis of 10hydroxytrilobacin, was synthesized as summarized in Scheme 4. *Cis*-THF aldehyde **2a** was obtained from allylsilane **1** via a known five-step sequence.⁸ The stereochemically mismatched⁸ [3 + 2]-annulation reaction of **2a** and allylsilane **3** mediated by SnCl₄ provided *cis/erythro/trans* bis-THF **25a** in 77% yield and with 15:1 diastereoselectivity. It was previously determined that the bulky TBS ether side chain in *cis*-THF aldehyde **2a** prevents the chelation with SnCl₄ in the stereochemically mismatched [3 + 2]-annulation reaction of **2a** with chiral, nonracemic allylsilanes homochirally related to **3**.⁸

We synthesized the three additional bis-THF diastereomers by simple modification of the conditions for the two [3 + 2]-annulation reactions summarized in Scheme 4. Use of an acetyl protecting group in **2b** allows chelation of **2b** and SnCl₄ in the stereochemically mismatched double asymmetric [3 + 2]-annulation reaction with allylsilane **3**,⁸ from which the *cis/threo/cis* bis-THF **25b** was obtained in 52% yield and with 20:1 diastereoselectivity.

Allylsilane **1** was also used in the modular synthesis of *trans*-THF **24b** by the chelate-controlled [3 + 2]-annulation with α -benzyloxyacetaldehyde, followed by protiodesilyla-

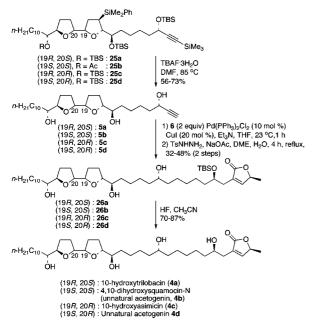
⁽²⁰⁾ The dehydration of **8** by treatment with MsCl and Et₃N was studied to determine if epimerization of the butenolide occurs under the mildly basic elimination conditions. As shown below, the dehydration of **8** via mesylate elimination proceeds without detectable racemization.



⁽¹⁶⁾ Nakajima, N.; Horita, K.; Abe, R.; Yonemitsu, O. *Tetrahedron Lett.* **1988**, *29*, 4139.

⁽¹⁷⁾ Yu, W.; Mei, Y.; Kang, Y.; Hua, Z.; Jin, Z. Org. Lett. 2004, 6, 3217.

⁽¹⁸⁾ Brown, H. C.; Jadhav, P. K. J. Am. Chem. Soc. 1983, 105, 2092.
(19) Evans, D. A.; Black, W. C. J. Am. Chem. Soc. 1993, 115, 4497.

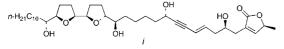


Scheme 5. Synthesis of 10-Hydroxytrilobacin and Three Diastereomeric Acetogenins

tion.²¹ Further elaboration of *trans*-THF **24b** to aldehyde **2c** followed by stereochemically matched double asymmetric [3 + 2]-annulation reactions with **3** led to the *trans/threo/trans* **(25c)** and the *trans/erythro/cis* **(25d)** bistetrahydro-furans via SnCl₄-mediated chelate-controlled and BF₃•OEt₂-mediated nonchelate-controlled annulations, respectively, both with > 20:1 dr.

With four diastereomeric bis-THF fragments (**25a**–**d**) in hand, their conversion to natural and unnatural acetogenins **4a**–**d** was completed (Scheme 5). First, bis-THF **25a**, the precursor to 10-hydroxytrilobacin, was subjected to protiodesilylation conditions (TBAF, DMF, 80–85 °C).²¹ These conditions effected removal of the *C*-phenyldimethylsilyl group as well as the three TBS ethers and the alkynyl silane and gave alkyne **5a** in 64% yield. Sonogashira coupling⁹ of alkyne **5a** and butenolide **6**²² was followed by enyne reduction using a large excess of TsNHNH₂ and NaOAc.²³ Finally, removal of the TBS ether of **26a** by treatment with

⁽²²⁾ Our original plan was to deprotect the C(4)-OTBS ether of **6** prior to the Sonogashira reaction with **5a**. This sequence provided *i* uneventfully (55% yield). However, reduction of the enyne unit of *i* provided 10-hydroxytrilobacin contaminated with an inseparable impurity. This problem required that we use the TBS ether protected vinyl iodide **6** in Scheme 5.



HF provided 10-hydroxy-trilobacin (4a). The enantiomeric purity of the butenolide unit of 4a was determined to be \geq 98% ee by NMR analysis on the bis-Mosher ester species derived from the 2',4-diol obtained by protection of 4a as the tetra-TBS ether followed by LiAlH₄ reduction.²⁴ The spectroscopic properties of synthetic 10-hydroxytrilobacin, as well as of the tetra-Mosher esters prepared from 4a, were in complete agreement with literature values.³ The longest linear sequence of this synthesis of 10-hydroxytrilobacin was 13 steps, originating from commercially available methyl-4-pentenoate (9).

The diastereomeric bis-THF fragments 25b-d were subjected to the same synthetic sequence as described for 25a, leading to the successful synthesis of three analogues of 10-hydroxytrilobacin: unnatural 4,10-dihydroxysquamocin-N (4b),²⁵ 10-hydroxyasimicin (4c),³ and the unnatural acetogenin 4d. Spectroscopic properties for 4c were in excellent agreement with literature values.³ Results of biological evaluation of these compounds will be reported elsewhere.

In conclusion, we have developed a convergent and highly stereoselective synthesis of 10-hydroxytrilobacin (4a). By utilizing nonchelate- and chelate-controlled [3 + 2]-annulation reaction conditions, we also achieved the modular synthesis of three diastereomers of 10-hydroxytrilobacin from the same precursors 1, 3, and 6. We anticipate that this strategy for synthesis of adjacent bis-THF acetogenins can be expanded to include use of the enantiomer 1, both enantiomers of the *syn* diastereomer of 1,²⁶ and allylsilane diastereomers of 3. This chemistry has the potential to provide a stereochemically diverse library of bis-THF acetogenins. Studies along these lines will be reported in due course.

Acknowledgment. This work was supported by the National Institutes of Health (GM038436)

Supporting Information Available: Experimental procedures and spectroscopic data for all new compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

OL801242D

(26) Lira, R.; Roush, W. R. Org. Lett. 2007, 9, 4315.

⁽²¹⁾ Heitzman, C. L.; Lambert, W. T.; Mertz, E.; Shotwell, J. B.; Tinsley, J. M.; Va, P.; Roush, W. R. *Org. Lett.* **2005**, *7*, 2405.

⁽²³⁾ Dewey, R. S.; van Tamelen, E. E. J. Am. Chem. Soc. **1961**, 83, 3729. (24) **4a** $\xrightarrow{1) \text{TBSCI, imidazole, DMF}}_{2) \text{ LAH, ether}}$ **4a** $\xrightarrow{2) \text{ LAH, ether}}_{3) \text{ MTPACI, pyridine, CH₂Cl₂}}$ **4b** $\xrightarrow{\text{CMTPA}}_{\text{CMTPA}}$ **4b** $\xrightarrow{2} \text{ BSO}}_{\text{CMTPA}}$

⁽²⁵⁾ References for the closely related natural product squamocin-N, see: (a) Sahai, M.; Singh, S.; Singh, M.; Gupta, Y. K.; Akashi, S.; Yuji, R.; Hirayama, K.; Asaki, H.; Araya, H.; Hara, N.; Eguchi, T.; Kakinuma, K.; Fujimoto, Y. *Chem. Pharm. Bull.* **1994**, *42*, 1163. (b) Araya, H. *Bull. Natl. Inst. Agro-Environ. Sci.* **2004**, *23*, 77.